

PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

EP.0050652



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ³: A23B 4/08, 4/14	A1	(11) International Publication Number: WO 81/03110 (43) International Publication Date: 12 November 1981 (12.11.81)
(21) International Application Number: PCT/US81/00524 (22) International Filing Date: 22 April 1981 (22.04.81) (31) Priority Application Numbers: 144,030 199,195 (32) Priority Dates: 28 April 1980 (28.04.80) 21 October 1980 (21.10.80) (33) Priority Country: US (71) Applicant; and (72) Inventor: BARTA, Kent, S. [US/US]; 601 Monroe, Jefferson City, MO 65101 (US). (74) Agent: LEAVITT, Donald, G.; Senniger, Powers, Leavitt and Roedel, 611 Olive Street, St. Louis, MO 63101 (US).		(81) Designated States: AT (European patent), AU, CH (European patent), DE (European patent), FR (European patent), GB (European patent), LU (European patent), NL (European patent), SE (European patent). Published <i>With international search report</i>
(54) Title: MEAT SANITIZING PROCESS (57) Abstract A process for preventing prespoilage proliferation of bacteria on fresh meat utilizes substantially sub-bactericidal concentrations of chlorine dioxide to suppress localized growth of such bacteria for up to at least 3 days post-slaughter. The chlorine dioxide solution is applied immediately post-slaughter and at remote times whenever renewed proliferation of such bacteria may occur. Formation of organic chlorine and oxidative by-product residuals is minimized upon treatment of meat at these chlorine dioxide concentrations.		

BEST AVAILABLE COPY

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	KP	Democratic People's Republic of Korea
AU	Australia	LI	Liechtenstein
BR	Brazil	LU	Luxembourg
CF	Central African Republic	MC	Monaco
CG	Congo	MG	Madagascar
CH	Switzerland	MW	Malawi
CM	Cameroon	NL	Netherlands
DE	Germany, Federal Republic of	NO	Norway
DK	Denmark	RO	Romania
FI	Finland	SE	Sweden
FR	France	SN	Senegal
GA	Gabon	SU	Soviet Union
GB	United Kingdom	TD	Chad
HU	Hungary	TG	Togo
JP	Japan	US	United States of America

MEAT SANITIZING PROCESS

Background of the Invention

Modern rapid distribution systems for fresh meats rely upon refrigeration and plant sanitation to provide a whole-
5 some product to the consumer. Although fresh meats in the United States are generally safe and free of hazardous levels of pathogens, the microbial quality may be poor; high levels of nonpathogenic spoilage bacteria, frequently present, dramatically shorten shelf life and affect the taste
10 and appearance of the meat. In response to this problem several states and the Federal Government have adopted or proposed standards regulating the bacterial content of fresh meat.

It is recognized that refrigeration is not a complete
15 solution to spoilage problems because the principal spoilage bacteria are psychrotrophes which grow well at 50°-15° C., or mesotrophes which are adapted to grow at lower temperatures. Ayres, J. C. (Food Research, 25:1, 1960) found that the dominant microorganisms growing on refrigerated
20 beef undergoing advanced spoilage comprised the Micrococci and Pseudomonads; even at temperatures as high as 15° C. the growing bacterial population was dominated by motile gram negative rods of the Pseudomonas generae.

These Pseudomonads, however, comprise only a minor
25 portion of the initial contaminating population. This population is extremely diverse, and species of Acinetobacter, Moraxella, Flavobacterium, and Aeromonas may be present in substantial numbers (See Jay & Shelef, Food Technol., 22:186 [1978]), in addition to members of the
30 Enterobacteriace. In the course of early spoilage at refrigerated temperatures, most of these species are displaced by nonproteolytic Pseudomonads. Off-odors characteristic of spoilage are detected when this sub-population



grows to a level in excess of 10^7 - 10^8 organism per square inch of meat surface.

The mechanism of displacement of some genera of bacteria by the Pseudomonads in the early stages of spoilage is not well understood. These organisms are physiologically versatile, and readily attack a wide variety of low molecular weight nitrogenous substances whose by-products of metabolism account for the off-odors associated with meat spoilage. However, it is known that attack of these substances is strongly repressed by glucose which comprises about 0.18 percent of the meat (Gill, C. O., J. Appl. Microbiol., 41:401 [1976]) Substantial development of the psychrotrophic Pseudomonads, Aeromonas, etc. in early spoilage may thus be preceded by the growth of other organisms which preferentially deplete surface glucose thereby relieving repression of the Pseudomonad development. As the temperature of chilling meat becomes refractory to bacterial growth generally, the Pseudomonads, etc. then emerge as the dominant population.

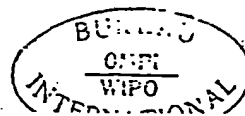
Bacterial contamination of meat has been the subject of extensive studies whose conclusions are here set forth. The microorganisms present in retail portions are derived principally from the initial bacterial load on the carcass surface immediately post-slaughter; thus, meat portions, such as hamburger, having high bacterial counts are traceable to carcasses having high surface contamination. (For example, see Elliot, et al., Appl. Microbiol., 9:452 [1961].) The primary source of such contamination is the gut and hide of the animal itself, although packing house environment (floors, chill room, cutting room, etc.), and handling by packing house workers are all substantial sources of contamination. (Frazier, Food Microbiology, Chapt. 16, 2ed., 1967.)



Contamination of carcasses from these sources is not uniform; initial bacterial counts vary from 10^2 organisms per square inch, or less, to greater than 10^6 organisms per square inch (Ingram & Roberts, "The Microbiology of the Red Meat Carcass and the Slaughterhouse," Meat Research Institute publication, Langford, Bristol, England, 1976). This variation in contamination occurs between different zones of the same carcass (including adjacent areas as small as one square inch), and among different carcasses. The relation of the magnitude and composition of initial contamination at any selected site to subsequent bacterial development at the same site has not heretofore been studied. Little is known about the kinetics of pre-spoilage proliferation of naturally-occurring mixed populations of such bacteria.

Several available processes eliminate bacteria from meat by killing them with a contact disinfectant(s) applied in the form of a spray to the carcass surface during chilling. U.S. Patent No. 3,745,026 (Hansen) discloses such a process utilizing 50-200 ppm of aqueous chlorine (hypochlorous acid). U.S. Patent No. 4,021,585 (Svoboda, et al.) describes an alternative process utilizing 5-50 ppm of aqueous chlorine dioxide. Both processes achieve reduced bacterial counts during the chill cycle (18-24 hrs. post-slaughter) by killing bacteria introduced onto the carcass during slaughter procedures. This is shown by the reduction in viable colony-forming bacteria present at the end of the chill cycle as compared with counts at the beginning of such period after carcasses are conveyed to the chill room from the kill floor.

A major limitation of processes utilizing aqueous chlorine (hypochlorous acid and its calcium or sodium salts) is rapid regrowth of microorganisms on the treated



surfaces. Several investigators have reported initial bactericidal action resulting in up to about 3 logs₁₀ of reduction in aerobic bacterial counts (Kotula, et al., J. Anim. Sci., 39:674 [1974]; Bailey, C., "Spray washing of lamb carcasses," 17th European meeting of meat researchers, Bristol, England [1971].) However, others have noted substantial bacterial regrowth during the first 48 hours post-slaughter; rapid regrowth accounts for net increases after initial reductions. (For example, see Anderson, et al., J. Food Sci., 42:326 [1977].)

A second major problem with such use of chlorinated contact disinfectants is reaction of the agent with meat components to produce chloro-organic derivatives such as chloro-substituted lipids, and oxidative products. These chemical derivatives may pose a health hazard, especially the class of halomethanes (known to be carcinogenic) formed by reaction of bactericidal levels of hypochlorous acid with humic or other organic substances. Reaction of chlorine dioxide at bactericidal concentrations with meat components results in low but detectable levels of organic chlorine, as noted in Cunningham & Lawrence (J. Assoc. Off. Anal. Chem., 62:482 [1979]).

Other agents such as inorganic and organic acids have also been applied to carcass surfaces in the form of aqueous sprays, as described in Carpenter, J. A., Proc. Meat Indust. Res. Conf., Chicago, 1972. Use of these agents has not received widespread acceptance because of reported surface damage to the carcass and off-odors and flavors imparted to the meat at bacteriostatic concentrations.

30 Summary of the Invention

A principal object of the present invention is to provide a process which suppresses prespoilage proliferation



of microorganisms on freshly slaughtered meat surfaces. A further object is to disable genera of bacteria whose transient growth on meat may promote development of the dominant spoilage populations without completely destroying meat microflora. A still further object is to provide a process for bacteria control of fresh meat which minimizes by-products formed by chemical reaction between the treating agent and the carcass surface.

Briefly summarizing, and without limiting the scope hereof, I have discovered that application to meat surfaces of aqueous chlorine dioxide at concentrations too low to be effective as a bactericide nevertheless substantially suppresses prespoilage proliferation of microorganisms. Application of substantially sub-bactericidal levels of chlorine dioxide up to about 4.0 ppm is effective in suppressing growth of the dominant spoilage bacteria and other populations which may promote their proliferation, for at least 3 days after application. At these concentrations, lower levels of reaction by-products are formed than at bactericidal concentrations.

Description of the Preferred Embodiment

In the present method, aqueous solutions of chlorine dioxide, so weak as to be substantially sub-bactericidal, are applied to meat surfaces immediately post-slaughter, and intermittently during subsequent chilling.

Typically, the chlorine dioxide is generated on site with conventional apparatus and formed into solution with potable water at a concentration of 0.5 to 4.5 ppm (mg/l), preferably about 1.0 to 4.0 ppm, prior to application to meat. The solution so formed is applied to meat surfaces during chilling, as hereinafter more fully described. In addition, the solution may be applied as a carcass wash on the kill-floor upon dehiding.



Conventionally, sanitizing agents are applied as an intermittent spray during chilling as disclosed in U.S. patent No. 4,021,585 (Svoboda, et al.). However, any method of applying the chlorine dioxide solution will be effective provided there is substantial contact between fresh solution and the meat during the course of 18 to 24 hour chill post-slaughter. Such alternative methods include, but are not limited to, dipping or misting continuously into the atmosphere at a constant low rate. Treatment of meat with 0.5 to 4.0 ppm chlorine dioxide is especially effective in avoiding proliferation of surface bacteria when applied intermittently for such intervals and in such volume as prescribed by Interim Chlorination Guidelines of the U.S. Dept. of Agriculture. Further application of the chlorine dioxide solution may be instituted during prolonged storage at remote times post-slaughter to avoid renewed proliferation of bacteria, as may occur upon partial warming of the meat prior to or coincident with boning, breaking, packaging, or shipping operations.

The animal carcass surface immediately post-slaughter and upon dehiding is contaminated by a diverse, mixed population of bacteria ranging in numbers from less than 100 organisms per square inch up to greater than 10^6 organisms per square inch. Applicant has discovered that an early event in the development of surface spoilage is an apparent sudden, rapid proliferation of organisms in one or more bacterial sub-populations following a latent period of approximately 24 hours during which growth is minimal. Such proliferation is herein termed a "bloom." For purposes of this application, a bloom is defined as an increase in bacterial numbers at any selected meat surface site two square inches in area, of at least $1.0 \log_{10}$, utilizing the assay as hereinafter more fully described in the Examples. A partial bloom is defined herein as 0.5 to $1.0 \log_{10}$ similar increase.



Applicant has further discovered that application to meat surfaces of about 0.5 to 4.0 ppm aqueous chlorine dioxide, as hereinabove set forth, is sufficient to substantially suppress blooms in spoilage genera of bacteria and in other species which may promote or potentiate their development. Such concentrations are substantially sub-bactericidal. In the meat slaughtering field, a bactericidal solution should be effective to kill at least about 85-90 percent of contaminating bacteria, as disclosed in U.S. Patent No. 4,021,585. That patent shows chlorine dioxide is so bactericidal at concentrations minimally 5.0 ppm.

In the present process, application of chlorine dioxide solutions at sub-bactericidal concentrations, that is, too low to achieve reliable 85-90 percent reductions in bacterial numbers, nevertheless suppresses bloom development which is now understood to be an early stage in spoilage. Significantly, such suppression is maintained for at least 3 days post-slaughter in contrast to sanitizing methods utilizing hypochlorous acid in which rapid regrowth is noted within 48 hours. Since the effect of chlorine dioxide is upon bacteria, whose taxonomic distribution is substantially identical on the surface of any species of fresh meat carcass, the present method is adaptable to the processing of fresh meat including, but not limited to, beef, pork, lamb and veal.

Reaction of hypochlorous acid (HOCl) or chlorine dioxide (ClO_2) with lipid components of meat may produce a great variety of chemical by-products, including chloro-substituted and oxidative derivatives. (For a comprehensive review, see W.J. Masschelein, Chlorine Dioxide: Chemistry and Environmental Impact of Oxychlorine Compounds. Ann Arbor Press: 1979.) Residues of these derivatives in meat may pose a potential health hazard to the consumer. Chlorinated by-products, thought to be formed



upon the chemical addition of inorganic chlorine to the double bonds of unsaturated fatty acids, are especially suspect because of their prolonged retention in the organs of test animals (Cunningham & Lawrence, Fd. Cosmet. Toxicol., 15:283 [1976]).

Studies of these chlorination reactions, as herein- after more fully described in the Examples, indicate that chlorine dioxide reacts very slowly with fatty acid double bonds compared to hypochlorous acid. In reaction with whole fat, chlorine dioxide reacts correspondingly more slowly than hypochlorous acid. The reaction appears to be nearly first order with respect to concentration. Use of chlorine dioxide at the sub-bactericidal levels of the present process will thus minimize formation of chemical by-product residues in meat treated therewith.

Other advantages of the present process will become apparent from the following examples.

EXAMPLE I

The following tests, utilizing a new bacteriological technique, demonstrate that chlorine dioxide applied to meat in a concentration of 0.5 to 4.5 ppm, and lower than heretofore used, effectively suppresses blooms in surface bacteria for at least a period of three days post-slaughter. Assay of bacterial numbers at selected carcass sites was performed as follows:

Sterile aluminum templates, each having a 2.0 square inch aperture were tag-pinned at randomly selected sites on beef carcass surfaces. Carcass halves were divided into chlorine dioxide treated and untreated control groups, with templates being affixed at corresponding positions on each treated and untreated carcass half.

At zero time, the carcass areas inscribed by each such template aperture were swabbed for 15 seconds with sterile polyester swab tips moistened in 0.1% peptone (Difco).



Swab tips were then individually agitated in 1.5 ml sterile 0.1% peptone to release adherent bacteria, and 1.0 ml aliquots of serial ten-fold dilutions in 0.1% peptone of this solution were plated by spreading on Plate Count Agar and MacConkey Agar. In these tests, zero time means prior to intermittent spray application during chill, but following an initial carcass wash with a solution of identical chlorine dioxide concentration.

After three days post-slaughter, the same template aperture sites were again swabbed, and swab specimens were diluted and plated as hereinabove described. Plated bacteria were incubated two to three days at room temperature to permit countable colonies to develop. The number of bacteria present in swabs, etc. is herein equated with colony forming units (CFU) arising on agar plates.

It will be apparent that the first swab at a particular site will remove some bacteria, but leave residual bacteria, which in turn are partially removed upon second swabbing. The efficiency of the swab procedure was determined as follows: sterile aluminum templates were affixed to carcass surfaces and their apertures swabbed as hereinabove described. Immediately thereupon, the outer 5.0 mm of carcass tissue inscribed by template apertures was aseptically excised, and homogenized for 2 minutes in 25.0 ml sterile 0.1% peptone. Serial dilutions of the homogenate in 0.1% peptone were plated on Plate Count Agar. Swab efficiency in percent is calculated by dividing the number of total bacteria (CFU) recovered from the swab by the sum of that number and the total number of bacteria present in the homogenate (CFU), and multiplying by 100. As empirically determined, average swab efficiencies were 38 percent; that is, 38 percent of initial contaminating bacteria were removed by the swab. Only a very few swab efficiency values were as low as 25 percent or as high as 75 percent.



In analyzing and interpreting results, let a equal the number of contaminating organisms actually present on any particular 2.0 square inch area of meat surface. Based on an average swab efficiency of 38 percent, the number of organisms removed on a first swab is $0.38 a$, leaving a residual population of $0.62 a$ bacteria. Assuming no growth or other increase in bacterial numbers over the interval between the first and second swab, a second swab will remove 38 percent of those bacteria remaining, or $0.38 (0.62 a)$ organisms. The apparent \log_{10} difference in bacterial counts recovered from the two swabs is: $\log_{10} 0.38 a - \log_{10} (0.38)(0.62 a) = -\log_{10} (0.38 a) / 0.38(0.62 a) = -0.2076$ logs. Any meat site having a non-growing bacterial population will thus show an apparent average $0.2076 \log_{10}$ decrease in bacterial counts.

Correspondingly, assume that following a first swab, a carcass site is treated with a bactericidal solution resulting minimally in an 80 percent reduction in bacterial numbers. This leaves 20 percent, or $0.2 (0.62 a)$ residual bacteria. A second swab will remove 38 percent of those bacteria remaining, or $(0.38)(0.2)((0.62 a)$ organisms. The apparent \log_{10} difference in bacterial counts recovered from the two swabs is: $\log_{10} .38 a - \log_{10} (0.38)(0.2)(0.62 a) = -\log_{10} .38 a / (0.38)(0.2)(0.62 a) = 0.9065$ logs. Any meat site upon which 80 percent of the bacteria are killed after removing the first swab sample will thus show an apparent average $.9065 \log_{10}$ decrease in bacterial counts.

A bloom in surface bacteria, as hereinabove set forth, is defined as $1.0 \log_{10}$ apparent increase in bacterial numbers, and a partial bloom as $0.5-1.0 \log_{10}$ such increase, utilizing this assay.

Table 1 summarizes representative data for carcass halves, divided into chlorine dioxide treated and untreated



TABLE 1A. Untreated Carcasses

	<u>Bacterial Counts*</u>		<u>Log₁₀ Change</u>	<u>Bloom**</u>
	<u>Zero</u>	<u>3 Days</u>		
1.	3.6532/2.3522	4.0530/3.1973	+0.3998/+0.8451	-/+
2.	4.8293/3.2945	5.5185/4.2833	+0.6892/+0.9888	+/+
3.	4.9685/3.8388	5.5740/4.7202	+0.6055/+0.8814	+/+
4.	4.4983/2.0792	5.4983/3.3222	+1.0000/+1.2430	+/+
5.	3.5740/2.0000	4.7324/3.1761	+1.1584/+1.1761	+/+
6.	4.4314/2.9542	5.2405/4.0845	+0.8091/+1.1304	+/+
7.	5.4983/3.7993	5.7201/3.9395	+0.2219/+0.1402	-/-
8.	4.2900/2.3802	5.3064/3.6230	+1.0164/+1.2430	+/+
9.	4.1399/2.0000	5.2659/3.1399	+1.1260/+1.1399	+/+
10.	4.8096/3.5011	4.8388/3.6532	+0.0292/+0.0621	-/-

B. Carcass Halves Treated with 0.8 ppm Chlorine Dioxide

15	1.	5.1055/3.2900	5.3064/3.9890	+0.2009/+0.6990	-/+
	2.	4.4456/2.8573	4.3404/2.0000	-0.1032/-0.8573	-/-
	3.	5.0952/4.1583	5.0334/3.0791	-0.0618/-1.0746	-/-
	4.	3.4983/2.5185	3.5434/2.6532	+0.0451/+0.1347	-/-

C. Carcass Halves Treated with 1.5 ppm Chlorine Dioxide

20	1.	2.5185/n.s.†	2.1761/n.s.	-0.3425/n.s.	-/-
	2.	4.9642/3.7993	4.8293/3.5911	-0.1248/-0.1972	-/-
	3.	3.1072/n.s.	2.5562/n.s.	-0.6409/n.s.	-/-
	4.	4.8751/3.6074	4.5378/2.8751	-0.3373/-0.7323	-/-

D. Carcass Halves Treated with 2.0 ppm Chlorine Dioxide

1.	5.4728/4.1055	4.7442/3.7993	-0.7285/-0.3061	-/-
2.	3.3936/n.s.	3.1538/n.s.	-0.2398/n.s.	-/-
3.	5.3522/3.0951	3.2659/2.0000	-2.0852/-1.1408	-/-
5 4.	3.3607/n.s.	3.0951/n.s.	-0.2655/n.s.	-/-

E. Carcass Halves Treated with 3.5 ppm Chlorine Dioxide

1.	4.6946/2.5740	4.7993/2.5563	+0.1047/-0.0169	-/-
2.	3.7202/n.s.	2.9754/n.s.	-0.7448/n.s.	-/-
3.	3.8573/2.6075	3.2900/2.4314	-0.5675/-0.1761	-/-
10 4.	4.1105/3.3522	4.1973/3.6675	+0.0867/+0.3152	-/-

F. Carcass Halves Treated with 4.5 ppm Chlorine Dioxide

1.	4.7993/n.s.	3.3434/n.s.	-1.4559/n.s.	-/-
2.	4.1941/3.1303	3.5670/2.0212	-0.6261/-0.1091	-/-
3.	5.2553/4.2589	3.4548/2.2553	-1.8005/-2.0036	-/-
15 4.	3.1972/n.s.	2.9004/n.s.	-0.2968/n.s.	-/-

*Bacterial counts in \log_{10} . Values to the left of the slash indicate bacterial counts on Plate Count Agar; numbers to the right of the slash indicate counts on MacConkey Agar.

20 **A plus sign indicates a bloom, that is, 1.0 \log_{10} or more increase in bacterial numbers; a plus/minus sign indicates a partial bloom, that is, 0.5-1.0 \log_{10} similar increase; a minus sign indicates no bloom, that is, less than 0.5 \log_{10} increase or net decrease in counts.

25

†n.s. means CFU too low to be significant.



groups, and assayed at zero time and at three days post-slaughter, according to the technique hereinabove set forth. Chlorine dioxide solutions were applied to treated carcass halves in a typical, conventional intermittent spray cycle. Spray was directed downwardly onto carcass halves from overhead spray nozzles in 1 minute pulses at 15 minute intervals during the first 6.5 hours of carcass chill, for a total exposure time of 26 minutes.

The data of Table 1 indicate that treatment of meat carcasses with aqueous chlorine dioxide in a concentration of 0.8 to 4.5 ppm effectively suppresses blooms in surface bacteria for at least three days post-slaughter. To validate this hypothesis, a chi-squared test was performed utilizing this data and other similar data (not shown) deemed redundant for illustrative purposes. Table 2 hereof summarizes this combined data.

TABLE 2

	<u>Number of</u> <u>Carcasses</u>	<u>Blooms</u>	<u>Partial</u> <u>Blooms</u>	<u>No Bloom</u>	<u>Total</u>
20	Untreated	19	8	12	= 39
	Treated	1	4	26	= 31
	<u>Total</u>	20	12	38	= 70

The chi-squared statistic, with two degrees of freedom, is 22.1, which is highly significant ($p < .005$). It is concluded that treatment of contaminated meat with 0.8 to 4.5 ppm chlorine dioxide profoundly reduces the likelihood of bloom development. Further statistical analysis comparing bloom suppression at various chlorine dioxide concentrations indicates substantial uniformity in effect within this concentration range.

Treatment of meat at a concentration of 0.5 ppm is



also effective in suppressing bacterial blooms; however, there are a higher number of partial blooms at this concentration than at 0.8 ppm. At concentrations substantially less than 0.5 ppm effectiveness rapidly diminishes, and
5 blooms may become more frequent than on untreated control carcasses because of increased water activity (A_w) at the meat surface. Although slight bacterial growth may occur at some carcass sites treated with 0.5 to 4.5 chlorine dioxide, such growth is significantly less than the increases
10 characteristic of prespoilage blooms and partial blooms.

In the foregoing tests utilizing 0.5 to 4.5 ppm chlorine dioxide, reliable 3 day reductions in bacterial counts were less than 0.9065 logs, the predicted minimum bactericidal value. Conventionally, however, bactericidal effi-
15 cacy is measured immediately after 20 to 24 hour chill instead of at 3 days post-slaughter. Corresponding control tests which demonstrate the sub-bactericidal effect of chlorine dioxide in the concentration range of the present process, are accordingly described hereinafter in Example
20 II.

In these tests, swab specimens were plated on Plate Count Agar and MacConkey Agar. Plate Count Agar is a nutritionally rich medium which supports the growth of most aerobic bacteria. MacConkey Agar contains bile salts and
25 crystal violet which inhibit the growth of Gram positive organisms and many other non-spoilage species; however, it is specifically permissive for Pseudomonas, Aeromonas, Flavobacterium, and most other Gram negative genera associated with meat spoilage. It is apparent from the data of
30 Table 1 hereinabove that spoilage organisms comprise a minor portion of the initial bacterial load on carcass surfaces. These data further indicate that an apparent bloom in total counts cannot always be accounted for solely by a

BIOLOGICAL
CEN
FBI

corresponding bloom in the spoilage classes of bacteria. However, the data of Parts B-F demonstrate that bloom suppression in all sub-populations of bacteria, and specifically in genera associated with spoilage, is achieved
5 utilizing sub-bactericidal concentrations of chlorine dioxide.

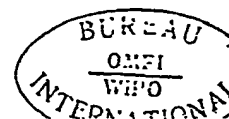
EXAMPLE II

Table 3 summarizes the results of tests identical in format to those of Example I hereof, except that a second
10 swab specimen was removed immediately at the end of the chill cycle approximately 20 hours post-slaughter.

The data of Part A of Table 3 shows that blooms develop on only about 10 percent of untreated carcass sites after 20 hours chill, compared to a corresponding 50 percent of such sites 3 days post-slaughter (See Table 2 hereinabove). Contaminating bacterial populations thus appear to have a substantial latent or lag period after deposition on meat surfaces. This is surprising because blooms in non-spoilage species as well as known psychrotrophes become
15 prevalent generally at 3 days, and long after carcasses have attained refrigerated temperatures.
20

Bactericidal reductions in the meat slaughtering field ideally should be upwardly 90 percent, as disclosed in U.S. Pat. No. 4,021,585 (Svoboda, et al.). The predicted apparent difference in total bacterial counts utilizing the
25 assay hereinabove described and corresponding to a 90 percent reduction in counts is -1.2076 logs, following the derivation of Example I: $\log_{10} .38 \underline{A}/(0.38)(0.1)(0.62 \underline{a}) = 1.2076$.

30 Referring to part E of Table 3, chlorine dioxide in a control concentration of 5.0 ppm results in reductions in



16

TABLE 3

		<u>Bacterial Counts*</u>		<u>Log₁₀ Change</u>	<u>Bloom</u>
		Zero	20 Hours		
<u>A. Untreated Carcasses</u>					
5	1.	3.0792/2.7993	3.4771/2.0414	+0.3999/-0.7579	-/-
	2.	4.0792/n.s. [†]	4.4149/n.s.	+0.3357/n.s.	-/-
	3.	4.7924/3.4472	5.0414/3.2304	+0.2490/-0.2168	-/-
	4.	5.0414/3.1139	5.0792/3.1463	-0.0378/-0.0324	-/-
	5.	3.8513/n.s.	4.2304/n.s.	+0.3791/n.s.	-/-
10	6.	3.7853/- [§]	4.9345/-	+1.1492/-	+/
	7.	3.9345/2.3010	4.3010/2.0414	+0.3665/-0.2596	-/-
	8.	4.4314/2.8451	4.7634/3.9638	+0.3320/+1.1187	-/+
<u>B. Carcass Halves Treated with 2.0 ppm Chlorine Dioxide</u>					
	1.	4.8388/3.7924	4.2304/2.7853	-0.6084/-1.0071	-/-
15	2.	4.7482/n.s.	4.4149/n.s.	-0.3333/n.s.	-/-
<u>C. Carcass Halves Treated with 3.0 ppm Chlorine Dioxide</u>					
	1.	4.0334/n.s.	4.1973/n.s.	+0.1639/n.s.	-/-
	2.	3.3664/n.s.	3.3802/n.s.	+0.0138/n.s.	-/-
	3.	3.7202/n.s.	3.8573/n.s.	+0.1371/n.s.	-/-
20	4.	3.8921/n.s.	3.7782/n.s.	-0.1139/n.s.	-/-
	5.	3.4548/n.s.	3.7559/n.s.	+0.3011/n.s.	-/-
	6.	3.3010/n.s.	3.7513/n.s.	+0.4503/n.s.	-/-
	7.	3.4149/-	2.6128/-	+0.8021/-	-/-



8. 5.1761/- 5.2195/- +0.0434/- -/

D. Carcass Halves Treated with 4.0 ppm Chlorine Dioxide

1.	3.4771/2.3522	3.7202/2.3010	+0.2431/-0.0512	-/-
2.	5.0952/2.4771	4.8482/2.5740	-0.2470/+0.0969	-/-
3.	4.4433/2.4314	4.3096/2.3522	-0.1337/-0.0792	-/-
4.	4.2175/2.3522	4.1303/2.4983	-0.0872/+0.1461	-/-
5.	3.7482/-	3.4471/-	-0.3011/-	-/-
6.	5.1461/2.7403	4.6628/2.6021	-0.4883/-0.1382	-/-

E. Carcass Halves Treated with 5.0 ppm Chlorine Dioxide

1.	3.3936/n.s.	2.6074/n.s.	-0.7862/n.s.	-/-
2.	4.0212/n.s.	2.5185/n.s.	-1.5027/n.s.	-/-
3.	4.9912/2.1761	3.9395/n.s.	-1.0517/-	-/-
4.	4.7559/2.9165	3.4314/2.7782	-1.3245/-0.1383	-/-
5.	3.7924/2.5563	2.6128/n.s.	-1.1796/-	-/-

*Bacterial counts in log₁₀; values to the left of the slash indicate bacterial counts on Plate Count Agar; values to the right of the slash indicate bacterial counts on MacConkey Agar.

**Plus sign indicates bloom, or 1.0 log₁₀ increase in bacterial numbers; minus sign indicates no bloom, or less than 0.15 log₁₀ increase in bacterial numbers.

1CFU too low to be significant.

SNot plated.



total bacterial counts (counts on Plate Count Agar) exceeding $1 \log_{10}$. An average of the values therein set forth is -1.1689 which closely approximates the predicted theoretical value of -1.2076 . It is concluded that 5.0 ppm chlorine dioxide is bactericidal in agreement with data disclosed in U.S. Pat. No. 4,021,585 (Svoboda, et al.).

However, the data of Parts B-D demonstrate that treatment of carcasses with 1.0 to 4.0 ppm chlorine dioxide fails to result in either minimally bactericidal reductions (-0.9065 logs) or ideal bactericidal reductions (-1.2076 logs), as determined immediately after chill.

Net changes in bacterial numbers at 20 hours are not appreciably different than at 3 days. This strongly suggests that chlorine dioxide at sub-bactericidal concentrations acts upon bacteria at a very early stage of spoilage prior to commencement of vegetative growth. These bacteria, while not killed, are disabled functionally, and their development on a meat substrate is aborted up to at least 3 days after the treating agent is dissipated. Their presence, however, may have important prophylactic value in preventing establishment of pathogens or new spoilage contaminants introduced subsequently.

EXAMPLE III

The following tests compare the rates of chlorine dioxide or hypochlorous acid reaction with fat alone or supplemented with linolenic acid, a long chain fatty acid containing three double bonds. These tests were performed according to the following general format simulating the two phase (liquid-solid) interaction of an aqueous chlorinating agent at the fat-containing surface of animal carcasses:



7.0 g of molten, whole, freshly rendered, beef fat were layered onto the bottom of each of several screw-capped 125 ml flasks, and allowed to solidify at room temperature. 15.0 ml of an aqueous solution containing various concentrations of hypochlorous acid (HOCl) or chlorine dioxide (ClO_2) were then individually pipetted onto the fat layers. The flasks were incubated at room temperature with gentle agitation (100 rpm) in a rotary shaker. At various times after the addition of the chlorinating agent, aliquots were withdrawn and assayed conventionally for the appropriate residual oxychlorine specie remaining in the supernatant solution. In these tests, disappearance of the chlorinating agent is equated with the formation of total reaction products plus the gaseous escape of the agent into the headspace of the flask.

Referring to Part A of Table 4, it is apparent that hypochlorous acid reacts more readily than chlorine dioxide with whole beef fat. Most of the observed decrease in HOCl concentration in the presence of fat (column 1, Part A) is attributable to chemical reaction with the fat layer, since only a small portion of the agent is flashed off into the headspace (see control in the absence of fat, column 5, Part A). In contrast, most of the observed decrease in chlorine dioxide concentration in the presence of fat (column 2, Part A) is attributable to flashing off of the agent (compare column 6 to column 2).

Ordinary animal fat contains no greater than approximately 2.0 percent unsaturated fatty acids having at least one double bond. Addition to the fat portion of 8 percent (w/w) linolenic acid (containing three double bonds) thus enriches the double bond content of the fat layer. Comparison of the data of column 1 with that of column 3 shows that double bond enrichment greatly enhances the reaction

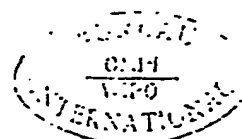


TABLE 4

A. Reaction of HOCl/ClO₂ with Fat Plus/Minus Linolenic Acid

Trial No.	1	2	3	4	5	6
5	Beef fat(7.0 g)	+	+	+	-	-
	Linolenic Acid(8%)	-	-	+	-	-
	HOCl (ppm)	300	-	300	300	-
	ClO ₂ (ppm)	-	325	-	-	325
	Concentration (ppm)					
	0 Time	300	325	300	325	300
10	2.0 Hour	120(40)*	190(58)	30(10)	180(55)	280(93)
	3.0 Hour	50(17)	175(54)	0(0)	160(49)	270(90)
						210(63)

B. Reaction of ClO₂ with Fat First Depleted by HOCl/ClO₂

Trial No.	1	2	3	4	5	6
15	Beef fat(7.0 g)	+	+	+	-	-
	First	HOCl	-	ClO ₂	-	-
	Second	ClO ₂	ClO ₂	HOCl	ClO ₂	HOCl
	Concentration (ppm)					
	0 Time	93	93	93	220	93
	1.5 Hour	88(95)	78(84)	93(100)	180(82)	93(100)
20	3.0 Hour	70(76)	57(62)	73(79)	170(78)	70(76)
	5.0 Hour	65(70)	40(44)	68(74)	150(69)	60(65)
						210(96)

C. Reaction of ClO₂ with Fat

Trial No.	1	2
Initial ClO ₂ Conc.	275**	85**
0.5 Hour	170(62)	55(65)
1.0 Hour	150(55)	40(48)
1.5 Hour	110(40)	35(42)
2.5 Hour	95(35)	27(32)
3.5 Hour	65(24)	18(22)

*Values in parantheses equal percent of starting concentration.

10 **Average of values obtained from two independent experiments.



rate of hypochlorous acid. In contrast, the rate of reaction between chlorine dioxide and enriched fat is not appreciably increased over the rate observed for fat alone (comparison of column 2 with column 4). It is concluded
5 that chlorine dioxide reacts very slowly with fatty acid double bonds compared to hypochlorous acid.

Referring to Part B of Table 4, the procedure of Part A was modified as follows: Layered fat was first exposed to saturating amounts of hypochlorous acid or chlorine di-
10 oxide by reacting the fat with an excess of the respective agent. The flask was then rinsed thoroughly with distilled water, and a second solution (15 ml) containing the same or different chlorinating agent was introduced. The concentration of the second such solution was monitored over a
15 time course (as in the experiment of Part A). The results indicate that pre-reacting the fat with excess hypochlorous acid effectively inhibits further reaction by chlorine dioxide (compare data of column 1 with control, column 5, and
20 column 3, fat not pre-reacted). This suggests that hypochlorous acid attacks substantially all of the fat substrate moieties otherwise available to chlorine dioxide, thereby depleting them.

Conversely, pre-reacting fat with chlorine dioxide fails to prevent further substantial reaction of the fat
25 with hypochlorous acid (compare data of column 4 with control, column 6). The results of Part A suggest that this additional reactivity may be due at least in part to the greater affinity of hypochlorous acid to fatty acid double bonds.

30 Referring to Part C of Table 4, the rate of reaction of chlorine dioxide in simple reaction with fat appears to be substantially independent of initial concentration. The rate of disappearance of the agent is approximately the



same at 275 ppm as at 85 ppm, as shown by comparing the percent remaining agent at various times. This suggests that reaction kinetics is very nearly first order with respect to concentration, indicating that the levels of reaction products formed in unit time are linearly proportional to concentration. Levels of total reaction products formed at sub-bactericidal concentrations of chlorine dioxide will be less than the levels formed at higher bactericidal concentrations. Use of chlorine dioxide at sub-bactericidal concentrations, and instead of hypochlorous acid, will thus minimize residues of organic reaction by-products in freshly slaughtered sanitized meat.



Claims

1. The process of avoiding proliferation of surface bacteria on freshly slaughtered meat comprising the steps of forming an aqueous solution of chlorine dioxide in a concentration of 0.5 to 4.0 ppm, said concentration being
5 great enough to substantially suppress bloom development of such bacteria on said meat, and less than that which is bactericidal to such bacteria, and applying said solution to said meat during chilling.

2. The process of avoiding proliferation of surface bacteria on meat carcasses and portions thereof comprising the steps of forming an aqueous solution of chlorine di-
oxide in a concentration of 0.5 to 4.0 ppm, said concentra-
5 tion being great enough to substantially suppress bloom development of such bacteria on said meat carcasses, and less than that which is bactericidal to such bacteria, and applying said solution to said meat during prolonged storage, whereby to suppress bloom development of such bacteria
10 at remote times post-slaughter.



FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

- | | | |
|---|--|-----|
| A | N, JOURNAL OF FOOD PROTECTION, ISSUED APRIL 1979, VOLUME 42, NUMBER 4, FIRSTENBERG-EDEN ET AL, SCANNING ELECTRON MICROSCOPIC INVESTIGATIONS INTO ATTACHMENT OF BACTERIA TO TEATS OF COWS, SEE PAGES 305-309. | 1-2 |
| A | N, JOURNAL OF MILK FOOD TECHNOLOGY, ISSUED NOVEMBER 1976, VOLUME 39, NUMBER 11, MAXEY, EVALUATION OF COLONY GROWTH OF BACTERIA AND THE SURFACE ENVIRONMENT OF BEEF, SEE PAGES 754-758. | 1-2 |

V. ☐ OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE ¹⁰

This international search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. ☐ Claim numbers _____, because they relate to subject matter ¹² not required to be searched by this Authority, namely:
2. ☐ Claim numbers _____, because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out ¹³, specifically:

VI. ☐ OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING ¹¹

This international Searching Authority found multiple inventions in this international application as follows:

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.
2. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:
3. ☐ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers:

Remark on Protest

- ☐ The additional search fees were accompanied by applicant's protest.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US81/00524

I. CLASSIFICATION OF SUBJECT MATTER (If several classification symbols apply, indicate all) ³		
According to International Patent Classification (IPC) or to both National Classification and IPC INT. CL. A23B 4/08, 14 U.S. CL. 426/332, 335, 532, 641, 652		
II. FIELDS SEARCHED		
Minimum Documentation Searched ⁴		
Classification System	Classification Symbols	
U.S.	426/92, 304, 310, 327, 332, 335, 532, 641, 652; 422/32, 37, 40	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ⁴		
III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴		
Category ⁶	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
A	US, A, 1,681,009, PUBLISHED 14 AUGUST 1928, PETERSEN.	1-2
A	US, A, 3,745,026, PUBLISHED 10 JULY 1973, HANSEN ET AL.	1-2
A	US, A, 3,819,329, PUBLISHED 25 JUNE 1974, KAESTNER ET AL.	1-2
A	US, A, 3,958,020, PUBLISHED 18 MAY 1976, DE VRIES.	1-2
A	US, A, 3,996,386, PUBLISHED 07 DECEMBER 1976, MALKKI ET AL.	1-2
A	US, A, 4,021,585, PUBLISHED 03 MAY 1977, SVOBODA ET AL.	1-2
A	N, JOURNAL OF FOOD SCIENCE, ISSUED 1979, VOLUME 44, LILLARD, LEVELS OF CHLORINE AND CHLORINE DIOXIDE OF EQUIVALENT BACTERICIDAL EFFECT IN POULTRY PRO- CESSING WATER, SEE PAGES 1594-1597.	1-2
A	N, JOURNAL OF FOOD PROTECTION, ISSUED MAY 1979, VOLUME 42, NUMBER 5, ANDERSON ET AL, MICROBIAL GROWTH ON PLATE BEEF DURING EXTENDED STORAGE DURING WASHING AND SANITIZING, SEE PAGES 389-392.	1-2
A	N, JOURNAL OF ANIMAL SCIENCE, ISSUED 1976, VOLUME 42, NUMBER 6, EMSWILER ET AL, BACTERICIDAL EFFECTIVENESS OF THREE CHLORINE SOURCES USED IN BEEF CARCASS WASHING, SEE PAGES 1445-1450.	1-2
* Special categories of cited documents: ¹⁵ <div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>"A" document defining the general state of the art</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document cited for special reason other than those referred to in the other categories</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> </div> <div style="width: 45%;"> <p>"P" document published prior to the international filing date but on or after the priority date claimed</p> <p>"T" later document published on or after the international filing date or priority date and not in conflict with the application, but cited to understand the principles or theory underlying the invention</p> <p>"X" document of particular relevance</p> </div> </div>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search ²		Date of Mailing of this International Search Report ²
29 JULY 1981		10 AUG 1981
International Searching Authority ¹		Signature of Authorized Officer ¹⁹
ISA/US		ARTHUR L. CORBIN

**This Page is Inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record**

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

☐ BLACK BORDERS

☐ IMAGE CUT OFF AT TOP, BOTTOM OR SIDES

☐ ~~FADED~~ TEXT OR DRAWING

☒ BLURRED OR ILLEGIBLE TEXT OR DRAWING

☐ SKEWED/SLANTED IMAGES

☐ COLOR OR BLACK AND WHITE PHOTOGRAPHS

☐ ~~GRAY~~ SCALE DOCUMENTS

☒ LINES OR MARKS ON ORIGINAL DOCUMENT

☐ REFERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY

☐ OTHER: _____

IMAGES ARE BEST AVAILABLE COPY.

As rescanning these documents will not correct the image problems checked, please do not report these problems to the IFW Image Problem Mailbox.

THIS PAGE BLANK (USPTO)